



Optimization of Ultrasound-Assisted Extraction of Phenolic Antioxidants from *Tectona grandis* Leaves, Using Experimental Design

Emmanuel N. Koffi^{1,2*}, Ibrahim Cissé³, Amian B. B. Kassi⁴, Paul R. Lozano⁵,
Augustin A. Adima³, Emmanuel N. Assidjo³ and Yves-Alain Bekro²

¹Higher Normal School of Abidjan, Department of Sciences and Technologies, 08 Bp 10 Abidjan 08, Côte d'Ivoire.

²Laboratory of Bioorganic Chemistry and Natural Substances, Nangui Abrogoua University, 02 BP 801 Abidjan 02, Côte d'Ivoire.

³Laboratory of Water Chemistry and Natural Substances, Felix Houphouët-Boigny National Polytechnic Institute, BP 1093 Yamoussoukro, Côte d'Ivoire.

⁴Laboratory of Organic Chemistry and Natural Substances, Felix Houphouët-Boigny University, 22 BP 582 Abidjan 22, Côte d'Ivoire.

⁵CIRAD, UMR-116 ISEM, TA B 16, 73 rue J. F. Breton, 34398 Montpellier cedex 5, France.

Authors' contributions

This work was carried out in collaboration between all authors. Author ENK designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript and managed literature searches. Authors IC, ABBK, PRL, AAA, ENA and YAB managed the manuscript and managed literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/EJMP/2015/20338

Editor(s):

(1) Marcello Iriti, Professor of Plant Biology and Pathology, Department of Agricultural and Environmental Sciences, Milan State University, Italy.

Reviewers:

(1) K. Yaprak Kantoğlu, TAEK Saraykoy Nuclear Research and Training Center, Turkey.

(2) Ibrahim Bulduk, Usak University, Turkey.

(3) Anonymous, The Research Institute of Forestry, China.

Complete Peer review History: <http://sciencedomain.org/review-history/11276>

Original Research Article

Received 22nd July 2015
Accepted 14th August 2015
Published 5th September 2015

ABSTRACT

Aims: This study aims to apply central composite design to optimize ultrasound-assisted extraction conditions in order to maximize simultaneously total polyphenols and antioxidant activity from aqueous leaves extract of *Tectona grandis*.

Study Design: Young teak leaves were collected in June 2011 from teak plantations in the center

*Corresponding author: E-mail: emmanuelkoffi@gmail.com;

of Côte d'Ivoire. After harvesting, the dried leaves were packed and shipped to CIRAD laboratory (Montpellier, France), where they were stored until processed and analyzed.

Place and Duration of Study: This study was carried out during season 2011-2012 in the CIRAD laboratory (Montpellier, France).

Methodology: Central composite design was used to simultaneously maximize ultrasound-assisted extraction of total polyphenols and antioxidant activity from *Tectona grandis* leaves. The design independent variables selected for this study were vegetal to liquid ratio (X_1 , leaves: citric acid; w:v), extraction time (X_2 , min) and solvent concentration (X_3 , citric acid concentration).

Results: Optimal condition obtained includes 10^{-2} N citric acid concentration, 16.25 g/L vegetal to citric acid ratio and 37.5 min for extraction. Under the above-mentioned condition, the experimental content of total polyphenols and antioxidant activity from aqueous leaves extract of *T. grandis* were $1,310 \mu\text{mol.g}^{-1}$ GAE and $431 \mu\text{mol.g}^{-1}$ TE, respectively. These results were well matched with their predicted values which are $1,300 \mu\text{mol.g}^{-1}$ GAE and $429 \mu\text{mol.g}^{-1}$ TE for polyphenols and antioxidant activity, respectively.

Conclusion: Ultrasound-assisted extraction was successfully optimized using central composite design to obtain an aqueous leaves extract of *Tectona grandis*, with optimized polyphenol content and antioxidant activity. Results indicated that this extraction method is a promising technique for extraction of phenolic antioxidants from *T. grandis*, as compared as infusion and decoction, and aqueous extracts of *T. grandis* leaves could be explored as a potential antioxidant agent for use in medicine against cardiovascular and cancer diseases.

Keywords: *Tectona grandis*; ultrasound-assisted extraction; optimization; central composite design; polyphenols; antioxidant activity.

1. INTRODUCTION

Tectona grandis Linn (common name: teak; Family: Verbenaceae), is grown in Côte d'Ivoire for its wood which have high commercial value. Different parts of this plant are also used by rural communities to prepare aqueous extracts used as traditional medicinal beverage [1,2]. Furthermore, several pharmacological activities have been attributed to *T. grandis* extracts, mainly antidiabetic activity [3,4] and antioxidant activity [5-8]. Antioxidants present in extracts of *T. grandis* could play an important role in the defense of human body against cardiovascular, aging and cancer diseases [9]. Previous studies showed that antioxidant activity exercise by plant extracts was due to their bioactive constituents such as polyphenols [10-12]. Polyphenols are also reported to have other pharmacologic properties such as anti-ulcer, anti-carcinogenic, anti-mutagenic activities, antibacterial, antiviral, anti-inflammatory, estrogenic, and anti-estrogenic properties [13,14]. To extract these compounds, Ivoirians used decoction and infusion. These extraction methods are time-consuming and thermally unsafe and the analysis of numerous constituents in plant material is limited by the extraction step [15]. For this reason, modern extraction techniques were described as alternative techniques to accelerate the extraction process. These modern techniques include: Supercritical fluid extraction, pressurized liquid extraction, microwave-assisted

extraction and ultrasound-assisted extraction [16]. The applications of these technologies, particularly ultrasound-assisted extraction offers many advantages, including temperature reduction and extraction time, which are very useful for the extraction of thermo labile and unstable compounds [17,18].

This study aims to apply response surface methodology with central composite design to optimize ultrasound-assisted extraction conditions such as vegetal to liquid ratio, extraction time and solvent concentration, in order to maximize simultaneously total polyphenol contents and antioxidant activity from *T. grandis* leaves. This experimental design provides high prediction (linear, quadratic, and interaction effects) of a response surface over the entire design space as compared as other experimental design [19].

2. MATERIALS AND METHODS

2.1 Plant Material

Entire young teak leaves (purple leaves) were collected from teak plantations (age ≤ 5 years) in the center of Côte d'Ivoire around Yamoussoukro area; then cut in strips of 10 cm of wide. After harvesting, the leaves were brought to LAPISEN laboratory (Yamoussoukro, Côte d'Ivoire) for drying at an average temperature of 30°C during daytime, and kept away from direct sun exposure

under an open-sided shed. Dried leaves were packed in plastic bags and shipped to CIRAD laboratory (Montpellier, France), where they were stored at 4°C until processed and analyzed.

2.2 Chemicals

All reagents were of analytical grade. Sodium carbonate salt (Na_2CO_3), monohydrated citric acid, dihydrated monosodium phosphate ($\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$), disodium hydrogen phosphate (Na_2HPO_4), Folin-Ciocalteu's reagent were purchased from Carlo Erba (Spain). Galic acid, trolox (6-hydroxy-2,5,7,8 tetramethylchroman-1-carboxylic acid), fluorescein, AAPH (2,2'-azobis (2-methylpropanimidamide) dihydrochloride) were purchased from Sigma-Aldrich (Germany).

2.3 Ultrasound-assisted Extraction Procedure

Ultrasound-assisted extraction (UAE) was carried out according to Adjé et al. [20], with a PEX 3 Sonifier ultrasounds system (R.E.U.S., Contes, France) composed of an stainless steel jug having 23 cm × 13.7 cm internal dimensions with a maximal capacity of 3 L, and a transducer, in the base of jug, operating at a frequency of 25 kHz with maximum input power of 150 W. Double layered mantle allowed us to control the temperature of the medium by cooling/heating systems. Generator output power was 150 W while the power dissipated in the medium was about 60 W per kilogram, as measured by calorimetry.

UAE was performed when applying ultrasound to plant material (5 to 20 g) put into water or acidified water ($V = 1$ L) during 15 to 45 min.

2.4 Total Polyphenol Contents

Total polyphenol content was determined by colorimetry, using Folin-Ciocalteu's (F-C) method [21,22]. To 30 μL sample extract, 2.5 mL of diluted Folin-Ciocalteu's reagent (1/10) were added. After 2 min incubation in dark at room temperature, 2 mL of aqueous sodium carbonate ($75 \text{ g}\cdot\text{L}^{-1}$) were added. After slight stirring, mixture was put in a water bath at 50°C for 15 min then cooled down. The absorbance was

measured at $\lambda = 760 \text{ nm}$ using a UV-visible spectrophotometer (Jenway 6705, Barloworld Scientific SAS, France). Total polyphenol content was expressed as $\mu\text{mol GAE}$ (Gallic Acid Equivalent) per gram of dried leaves water-extracted. Samples were analyzed in triplicate.

2.5 Antioxidant Capacity

Antioxidant capacity was carried by Oxygen radical absorbance capacity (ORAC) assay, as described by Ou et al. [23]. The automated ORAC assay was carried out on a VICTOR™ X3 Multilabel Plate Reader (Perkin-Elmer, USA) with fluorescence filters for an excitation wavelength at 485 nm and an emission wavelength at 535 nm [24]. To start reaction, 100 μL of fluorescein ($78 \text{ nmol}\cdot\text{L}^{-1}$) and 100 μL of diluted sample, phosphate buffer (pH 7.4) or standard (Trolox $5\text{-}50 \mu\text{mol}\cdot\text{L}^{-1}$) were placed in each well of a 96 well-plate and pre-incubated during 15 min. After, 50 μL of AAPH ($221 \text{ mmol}\cdot\text{L}^{-1}$) were added into the wells. Fluorescence was measured every minute during 60 min with emission and excitation wavelength set at 485 and 535 nm, respectively, at 37°C. ORAC values were calculated as area under the curve (AUC) and were expressed as $\mu\text{mol TE}$ (Trolox Equivalent) per gram of dried leaves water-extracted. Samples were analyzed in triplicate.

2.6 Central Composite Experimental Design

A five level, three variable central composite designs was applied to determine the best combination of extraction variables for the extraction of phenolic compounds and antioxidants from *T. grandis* leaves. Central composite design comprised 20 experimental runs with eight factorial points, six axial points (two axial points on each design variable axis at a distance of 1.68 from the design center) and six replicates at the center point. Design independent variables selected for this study were the vegetal to liquid ratio (X_1 , leaves: citric acid; w:v), extraction time (X_2 , min) and solvent concentration (X_3 , citric acid concentration) (Table 1).

Table 1. Independent variables and their coded and actual values used for optimization

Independent variable	Symbol	Coded level				
		-1.68	-1	0	1	1.68
Vegetal: liquid ratio (g/L)	X_1	5	8	12.5	18	20
Extraction time (min)	X_2	15	21	30	39	45
Citric acid conc. (10^{-3} N)	X_3	0	3.5	5	6.5	10

Two experimental responses were studied (total polyphenol contents (Y_1) and antioxidant capacity (Y_2)). Citric acid volume used was set to 1 L, during all experiments.

2.7 Statistical analysis

Multiple linear regression analysis was performed using the *Statistica 8* software (Stat Soft, Inc., USA). Experimental data were fitted to the following second-order polynomial model and regression coefficients were obtained.

$$Y_n = b_0 + b_1X_1 + b_2X_2 + b_3X_3 + b_{11}X_1^2 + b_{22}X_2^2 + b_{33}X_3^2 + b_{12}X_1X_2 + b_{13}X_1X_3 + b_{23}X_2X_3 \quad (1)$$

Where Y_n is the experimental response; X_1 , X_2 and X_3 correspond to the independent variables namely vegetal to liquid ratio, extraction time and citric acid concentration, respectively. The b_n values represent corresponding regression coefficients.

According to the experimental data, the fitting model represented by equation (1) was constructed and the statistical significance of the model terms was examined by regression analysis and analysis of variance (ANOVA).

3. RESULTS AND DISCUSSION

3.1 Experimental Responses Obtained using Central Composite Design

In order to optimize simultaneously of total polyphenol contents and antioxidant activity from *Tectona grandis* leaves under sonication, a central composite design was developed as represented in Table 2. This table presents also experimental values of total polyphenol contents and antioxidant activity from aqueous leaves extract of *T. grandis* at various conditions.

Good correlation ($R^2 = 0.86$) are observed between phenolic compounds and antioxidants extracted from aqueous leaves extracts of *T. grandis* (Fig. 1).

Similar correlation between polyphenol with antioxidant activity was observed by Cortés-Rojas et al.[25] and Lizcano et al. [26]. Therefore, aqueous leaf extract of *T. grandis* could be beneficial to human body. Indeed, natural antioxidants play an important role in the defense of human body against cancer and cardiovascular diseases [9].

Table 2. Response surface central composite design and experimental results

Test set	Independent variables			Experimental responses	
	X_1 (g/L)	X_2 (min)	X_3 (10^{-3} N)	Y_1 ($\mu\text{mol.g}^{-1}$ GAE)	Y_2 ($\mu\text{mol.g}^{-1}$ TE)
1	-1 (8)	-1 (21)	-1 (3.5)	421.45±10	163.27±1
2	1(18)	-1 (21)	-1 (3.5)	693.59±12	251.54±2
3	-1(8)	1 (39)	-1 (3.5)	935.28±7	316.15±5
4	1(18)	1 (39)	-1 (3.5)	1,145.68±15	323.12±3
5	-1(8)	-1 (21)	1 (6.5)	869.4±22	310.52±2
6	1(18)	-1 (21)	1 (6.5)	1,170.85±13	405.03±7
7	-1(8)	1 (39)	1 (6.5)	1,086.31±20	357.61±2
8	1(18)	1 (39)	1 (6.5)	1,276.92±18	391.49±2
9	-1.68 (5)	0 (30)	0 (5)	781.09±15	299.47±5
10	1.68 (20)	0 (30)	0 (5)	1,119.74±12	338.47±5
11	0 (12.5)	-1.68 (15)	0 (5)	592.55±10	157.01±2
12	0 (12.5)	1.68 (45)	0 (5)	1,212.88±13	366.14±3
13	0 (12.5)	0 (30)	-1.68 (0)	843.99±20	313.73±2
14	0 (12.5)	0 (30)	1.68 (10)	1,291.71±18	428.94±1
15	0 (12.5)	0 (30)	0 (5)	1,117.19±15	330.91±3
16	0 (12.5)	0 (30)	0 (5)	1,185.7±23	342.86±5
17	0 (12.5)	0 (30)	0 (5)	1,148.81±25	381.12±7
18	0 (12.5)	0 (30)	0 (5)	1,192.73±10	366.51±3
19	0 (12.5)	0 (30)	0 (5)	1,106.65±12	346.41±2
20	0 (12.5)	0 (30)	0 (5)	1,125.97±22	356.22±5

3.2 Fitting the Response Surface Models

By referring to Table 3, the coefficient of determination (R^2) value for regression model of polyphenols and antioxidant capacity were 0.99 and 0.93, respectively; which were closed to 1. These suggest that the predicted second order polynomial models defined well the real behavior of the system [27]. Their non-significant lack of fit also indicated that these models were good fit [28]. The lack-of-fit measured the failure of the model to represent data in the experimental domain at points which are not included in the regression [29].

3.3 Effect of Process Variables on Polyphenol Extraction

The polyphenols from *T. grandis* aqueous leaves extracts obtained by UAE based on central composite design are shown in Table 2. Multiple regression analysis was performed on the experimental data and the coefficients of the model are evaluated for significance. Extraction time and solvent concentration effects are highly significant ($P = .001$). Similar results were obtained when extracting phenolic compounds from grape [30]. Also, vegetal ratio played an important role in phenolic compounds extraction from plant material. Coefficients values for polyphenols as presented in Table 3 are used for a final predictive equation neglecting the non-significant terms as given below:

$$Y_1 = 1147 + 113 X_1 + 171 X_2 + 143 X_3 - 72 X_1^2 - 89 X_2^2 - 30 X_3^2 - 80 X_2 X_3 \quad (2)$$

All linear terms (X_1 , X_2 , and X_3), quadratic term (X_1^2 , X_2^2 and X_3^2) and interaction between X_2 and X_3 were significant. Those significant terms had a remarkable impact on polyphenols extraction from *T. grandis* leaves, whereas the non-significant terms had a negligible influence. The negative interaction between X_2 and X_3 indicate that interaction decrease phenolic compounds extracted when increasing both variables during their extraction.

In order to assess the effects of the extraction conditions on phenolic compounds extraction from *T. grandis* leaves, response surfaces and contour plots are constructed in accordance with equation (2). Fig. 2A shows the effects of vegetal to liquid ratio and extraction time on total phenolic contents from *T. grandis* leaves. It shows that lower level of both variables gives low amounts of polyphenols. However, total phenolic Content increases with vegetal to liquid ratio increases at a fixed extraction time, up to a certain limit. Beyond this limit, total polyphenol contents slightly decreases, which indicates that it greater content could be achieved if moderate vegetal to liquid ratio and extraction time were selected. This is due to significant negative quadratic effect of these variables (vegetal to liquid ratio and extraction time) on polyphenol extraction.

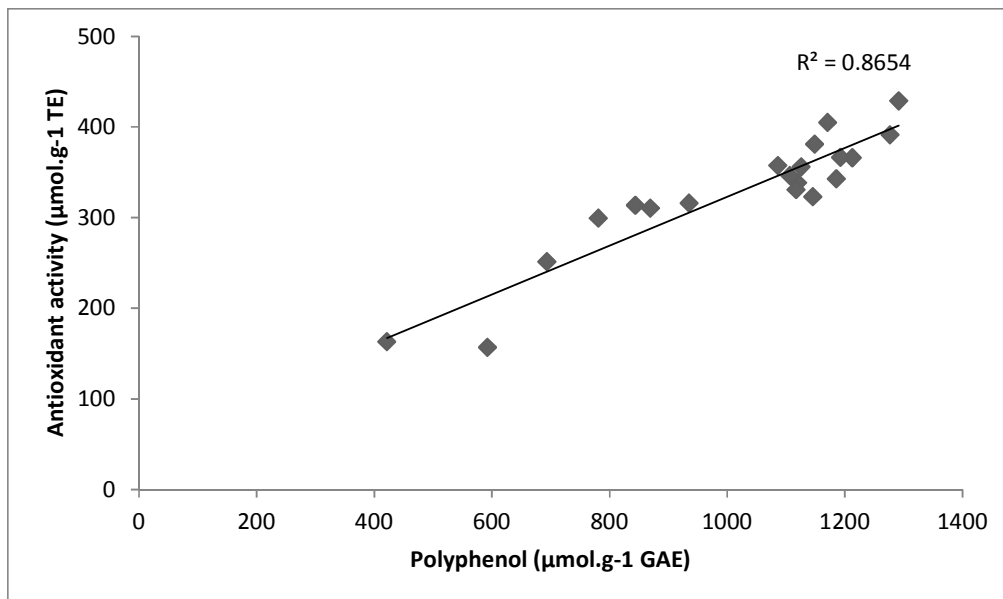


Fig. 1. Correlation between antioxidant capacity and polyphenol contents from *T. grandis* leaves

Table3. Regression coefficients of predicted quadratic polynomial models for polyphenols and antioxidant capacity

Coefficients	Coefficient estimated	
	Polyphenols	Antioxidant activity
b₀	1,146.60 ^{***}	354.01 ^{***}
Linear		
b₁	113.06 ^{***}	21.18 ^{**}
b₂	170.76 ^{***}	44.64 ^{***}
b₃	143.54 ^{***}	44.24 ^{***}
Quadratic		
b₁₁	-72.00 ^{***}	-12.43 [*]
b₂₂	-88.86 ^{***}	-32.72 ^{***}
b₃₃	-30.49 [*]	6.08 ^{ns}
Cross products		
b₁₂	-21.57 ^{ns}	-17.74 [*]
b₁₃	1.19 ^{ns}	4.14 ^{ns}
b₂₃	-80.37 [*]	-23.86 [*]
R²	0.99	0.93
Lack of fit (P-value)	0.60 ^{ns}	0.12 ^{ns}

^{*}Significant at $p = .05$; ^{**}Significant at $p = .01$; ^{***}Significant at $p = .001$; ns: no significant; R^2 : Regression coefficient; p : probability

In fact, a prolonged extraction time would allow all the plant cells to be completely cracked by acoustic cavitations, thus the extraction yield would increase within a certain time. On the other hand, completely ruptured plant cells would also allow various compounds such as insoluble and cytosolic solvent. In addition, target constituents might also be re-absorbed on smashed plants particles, thus affecting recovered compounds [18].

Interaction between vegetal to liquid ratio and citric acid concentration has positive effect on polyphenols extraction (Fig. 2B). Indeed, when increasing vegetal to liquid ratio and citric acid concentration, in experimental design, polyphenol contents increases.

Fig. 2C shows extraction time and citric acid concentration effects on total polyphenols extraction from *T. grandis* leaves at a constant vegetal to liquid ratio (16.25 g/L). Increase of both independent variables enhances polyphenols extraction. Polyphenol contents are particularly increased with prolonged extraction time at certain value. When total polyphenol extracted is maximal, extraction time extension would cause their slight degradation.

3.4 Effect of Process Variables on Antioxidant Extraction

The results show that antioxidant capacity from *Tectona grandis* aqueous leaf extracts, under different combinations, range from 163.27 to

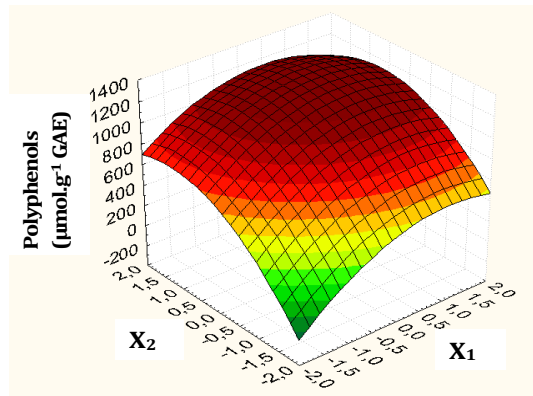
428.94 $\mu\text{mol TE/g}$ of leaves. By applying multiple regression analysis, relationship between tested independent variables and antioxidant extraction are explained in equation (3):

$$Y_3 = 354 + 21 X_1 + 45 X_2 + 44 X_3 - 12 X_1^2 - 33 X_2^2 - 18 X_1 X_2 - 24 X_2 X_3 \quad (3)$$

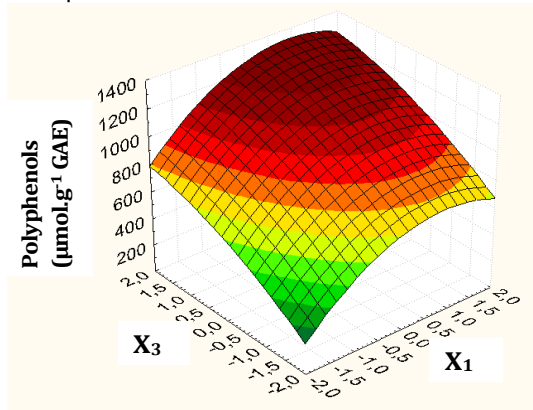
All linear terms (X_1 , X_2 , and X_3), quadratic term (X_1^2 and X_2^2) and interactions (between X_2 and X_3 ; between X_2 and X_3) are significant. Those significant terms had a remarkable impact on antioxidant extraction from *T. grandis* leaves. Extraction time and solvent concentration effects are highly significant ($P = .001$), on antioxidant extraction. Similar results were obtained when extracting total polyphenol from *T. grandis* leaves (Table 2). The surface plot in Fig. 3A shows vegetal to liquid ratio and extraction time effects on antioxidants extraction. Increasing of vegetal to liquid ratio doesn't significantly affect antioxidant extraction. But, extraction time are highly influenced antioxidant contents ($P = .001$).

Fig. 3B denotes citric acid concentration and vegetal to liquid ratio effects, on antioxidant extraction. When increasing citric acid concentration, antioxidant contents also increase.

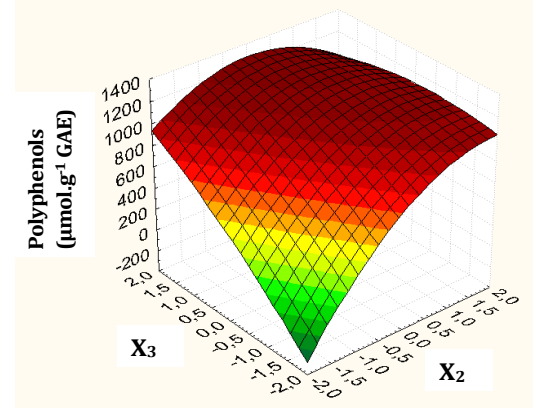
Fig. 3C shows citric acid concentration and extraction time effects on antioxidant from *T. grandis* leaves. Increase of both variables (acid concentration and extraction time on antioxidant) increases antioxidant extraction. Interaction between these variables is not significant (Table 2).



(A) : an effect of vegetal to liquid ratio and extraction time at citric acid concentration of 5.10-3N



(B) : a function of vegetal to liquid ratio and citric acid concentration at extraction time of 30 min



(C) : a function of extraction time and citric acid concentration at vegetal to liquid ratio of 12.5 g.L-1

Fig. 2. Response surface of phenolic contents of *Tectona grandis* leaves extract

3.5 Optimization and Experimental Validation

Optimal extraction condition with a citric acid concentration of 10^{-2} N, vegetal: citric acid ratio of 16.25 g/L and extraction time of 37.5 min are predicted using *Statistica 8.0* software desirability

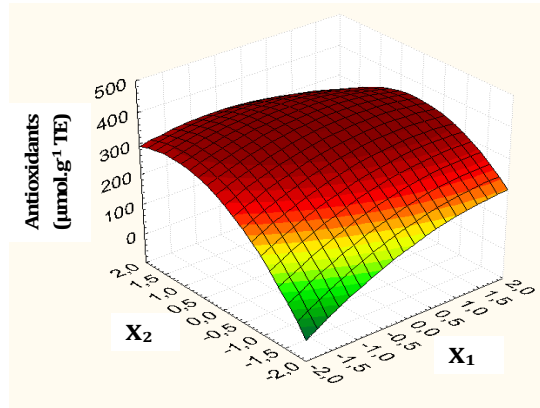
function, to simultaneously maximize polyphenols and antioxidant activity (Table 4).

Under the above-mentioned conditions, the experimental results are very close to predicted one. This implies that there are a high fit degree between observed values in experiment and predicted ones from the regression model [27].

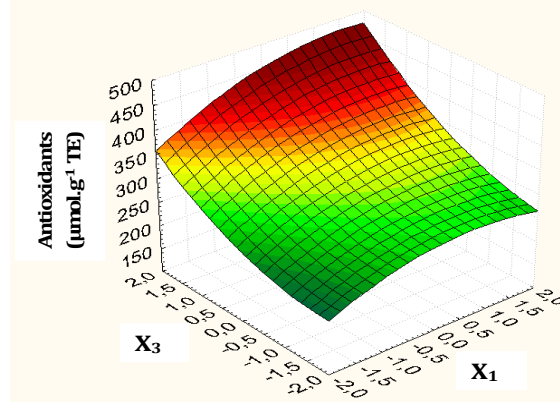
Table 4. Predicted and experimental values of responses under optimal condition and classic extraction

Compounds	UAE optimum condition		Classic extraction	
	Observed value	Predicted value	Infusion	Decoction
Polyphenols ($\mu\text{mol.g}^{-1}$ GAE)	$1,310 \pm 10^a$	$1,300^a$	$1,117 \pm 25^b$	$1,033 \pm 13^c$
Antioxidant activity ($\mu\text{mol.g}^{-1}$ TE)	431 ± 5^a	429^a	nd	nd

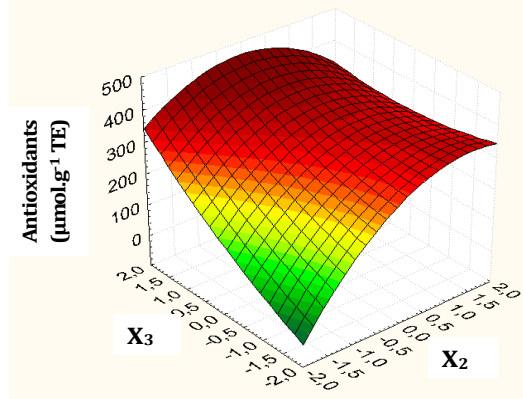
NB: in every line, the averages not followed by the same letter are significantly different at $p=0.05$



(A): an effect of vegetal to liquid ratio and extraction time at citric acid concentration of 5.10-3N



(B): a function of vegetal to liquid ratio and citric acid concentration at extraction time of 30 min



(C): a function of extraction time and citric acid concentration at vegetal to liquid ratio of 12.5 g.L-1

Fig. 3. Response surface of antioxidant contents of *Tectona grandis* leaves extract

The total polyphenol ($1,310 \pm 10 \mu\text{mol.g}^{-1}$ GAE) from aqueous leaves ultrasound-assisted optimized extract of *T. grandis* are higher than those of homemade extracts obtained with extraction time varying between 3-24h. Similar result was obtained by Koffiet et al. [31], when extracting phenolic compounds from leaves of *Justicia secunda*. So, ultrasound-assisted extraction gives a shorter extraction time and higher amount of phenolic compounds as compared to classic extraction by infusion and decoction.

4. CONCLUSION

In this study, the optimum condition of ultrasound-assisted extraction (UAE) of polyphenols and antioxidant activity from *T. grandis* was investigated. The optimal condition of UAE was achieved based on response surface methodology with Central Composite design. The optimal extraction condition with a citric acid concentration of 10^{-2} N, vegetal: citric acid ratio of 16.25 g/L and extraction time of 37.5 min was predicted using the desirability function. Under this optimal extraction conditions, polyphenol content and antioxidant activity were $1,310 \mu\text{mol.g}^{-1}$ EAG and $431 \mu\text{mol.g}^{-1}$ TE, respectively, which were well closed with the predicted value ($1,300 \mu\text{mol.g}^{-1}$ EAG and $429 \mu\text{mol.g}^{-1}$ TE, respectively). Results indicated that this extraction method is a promising technique for extraction of phenolic antioxidants from *T. grandis*, as compared as infusion and decoction, and aqueous extracts of *T. grandis* leaves could be explored as a potential antioxidant agent for use in medicine against cardiovascular and cancer diseases.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Tra Bi FH, Irie GM, N' Gaman K, Mahou CHB. Études de quelques plantes thérapeutiques utilisées dans le traitement de l' hypertension artérielle et du diabète: Deux maladies émergentes en Côte d'Ivoire. *Sci & Nat.* 2008;5(1):39-48.
2. Aradhana R, Rao KNV, Banji D, Chaithanya RK. A review on *Tectona grandis* linn: Chemistry and Medicinal uses (Family: Verbenaceae). *Herb. Tech Ind.* 2010;6-9.
3. Ghaisas MM, Navghare VV, Takawale AR, Zope VS, Phanse MA. Antidiabetic and nephroprotective effect of *Tectona grandis* linn. in alloxan induced diabetes. *ARS Pharm.* 2010;51(4):195-206.
4. Pooja VS, Samanta KC. Hypoglycemic activity of methanolic extract of *Tectona grandis* linn. Root in alloxan induced diabetic rats. *J. Appl. Pharm. Sci.* 2011; 1(4):106-109.
5. Naira N, Karvekar MD. Anti microbial and anti-oxidant properties of the isolated compounds from the methanolic extract from the leaves of *Tectona grandis*. *J. Basic Clin. Pharm.* 2011;2(4):163-165.
6. Shruthi DP, Sunith KE, Haritha KE, Govindappa M, Siddalingeshwara KG. Phytochemical screening, antioxidant and anti-inflammatory activity of different extracts from leaf, stem and bark of *Tectona grandis*. *Int J Res Pharmacol Pharmacother.* 2012;1(2):140-146.
7. Setiawan C, Purnomo H, Kusnadi J. Application of Microwave-Assisted Extraction on Teak (*Tectona grandis*) Leaves Antioxidant Extraction. *Res J Pharm Biol Chem Sci.* 2013;4(3):1012-1018.
8. Rao KNV, Aradhana R, Banji D, Chaitanya R, Kumar A.A. In-Vitro Anti-Oxidant and free radical scavenging activity of various extracts of *Tectona grandis* Linn. Leaves. *J. Pharm. Res.* 2011;4(2):440-442.
9. Oroian M, Escriche I. Antioxidants: Characterization, natural sources, extraction and analysis. *Food Res Int.* 2015;74:10-36.
10. Apak R, Güçlü K, Demirata B, Özyürek M, Çelik SE, Bektaşoğlu B, et al. Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. *Molecules.* 2007;12(7):1496-1547.
11. Dai J, Mumper RJ. Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules.* 2010;15(10):7313-7352.
12. Roleira FMF, Tavares-da-Silva EJ, Varela CL, Costa SC, Silva T, Garrido J, et al. Plant derived and dietary phenolic antioxidants: Anticancer properties. *Food Chem* 183. 2015;183:235-242.

13. Miyazawa M, Sakano K, Nakamura SI, Kosaka H. Antimutagenic activity of isoflavones from soybeans seeds (*Glycine max* Merrill.). J. Agric. Food Chem. 1999;47: 1346-1349.
14. Tapas AR, Sakarkar DM, Kakde RB. Flavonoids as nutraceuticals: A review. Tropical Journal of Pharmaceutical Research. 2008;7(3):1089-1099.
15. Mandal V, Mohan Y, Hemalatha S. Microwave assisted extraction-An innovative and promising extraction tool for medicinal plant research. Pharmacogn. Rev. 2007;1(1):7-18.
16. Garcia-Salas P, Morales-Soto A, Segura-Carretero A, Fernández-gutiérrez A. Phenolic-compound-extraction systems for fruit and vegetable samples. Molecules. 2010;15(12):8813-8826.
17. Shirsath SR, Sonawane SH, Gogate PR. Intensification of extraction of natural products using ultrasonic irradiations-A review of current status. Chem. Eng. Proces: Process Intensif. 2012;53:10-23.
18. Vilkhuk K, Mawson R, Simons L, Bates D. Applications and opportunities for ultrasound assisted extraction in the food industries: A review. Innov. Food Sci Emerg Technol. 2008;9:161-169.
19. Yoon J. Application of experimental design and optimization to PFC model calibration in uniaxial compression simulation. International Journal of Rock Mechanics and Mining Sciences. 2007;44(6):871-889.
20. Adjé F, Lozano YF, Lozano P, Adima A, Chemat F, Gaydou EM. Optimization of anthocyanin, flavonol and phenolic acid extractions from *Delonix regia* tree flowers using ultrasound-assisted water extraction. Ind Crops Prod. 2010;32(3):439-444.
21. Singleton VL, Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phospho-tungstic acid reagents. Am. J. Enol. Viticult. 1965;16:144-158.
22. Wood JE, Senthilmohan ST, Peskin AV. Antioxidant activity of procyanidin-containing plant extracts at different pHs. Food Chem. 2002;77(2):155-161.
23. Ou B, Hampsch-Woodill M, Prior RL. Development and validation of an improved oxygen radical absorbance capacity assay using fluorescein as the fluorescent probe. J. Agric. Food Chem. 2001;49(10):4619-4626.
24. Zulueta A, Esteve MJ, Frígola A. Orac and teac assays comparison to measure the antioxidant capacity of food products. Food Chem. 2009;114(1):310-316.
25. Cortés-Rojas DF, Souza CRF, Oliveira WP. Optimisation of the extraction of phenolic compounds and antioxidant activity from aerial parts of *Bidens pilosa* L. using response surface methodology. Int J Food Sci Technol. 2011;46(11):2420-2427.
26. Lizcano LJ, Bakkali F, Ruiz-Larrea MB, Ruiz-Sanz JI. Antioxidant activity and polyphenol content of aqueous extracts from Colombian Amazonian plants with medicinal use. Food Chem. 2010;119: 1566-1570.
27. Chan SW, Lee CY, Yap CF, Wan A, Ho CW. Optimisation of extraction conditions for phenolic compounds from limau purut (*Citrus hystrix*) peels. Int Food Res. J. 2009;16:203-213.
28. Silva EM, Souza JNS, Rogez H, Rees J.F, Larondelle Y. Antioxidant activities and polyphenolic contents of fifteen selected plant species from the Amazonian region. Food Chem. 2007;101(3):1012-1018.
29. Wang L, Wang G, Zhang J, Zhang G, Jia L, Liu X, et al. Extraction optimization and antioxidant activity of intracellular selenium polysaccharide by *Cordyceps sinensis* SU-02. Carbohydr. Polym. 2011;86(4):1745-1750.
30. Ghafoor K, Hui T, Choi YH. Optimisation of ultrasonic-assisted extraction of total anthocyanins from grape peel using response surface methodology. J. Food Biochem. 2009;35(3):735-746.
31. Koffi EN, Le Guernevé C, Lozano PR, Meudec E, Adjé FA, Bekro YA, et al. Polyphenol extraction and characterization of *Justicia secunda* Vahl leaves for traditional medicinal uses. Ind Crops Prod. 2013;49:682-689.

© 2015 Koffi et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
 The peer review history for this paper can be accessed here:
<http://sciencedomain.org/review-history/11276>